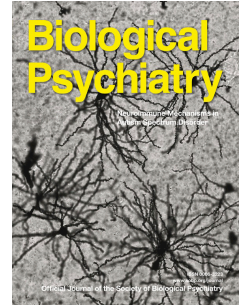


# Journal Pre-proof



Reduced neuronal cAMP in the **nucleus accumbens** damages blood-brain barrier integrity and promotes stress vulnerability

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2

3 **Abstract:**

4 **Background:** Studies have suggested that chronic social stress specifically downregulates  
5 endothelial tight junction protein expression in the nucleus accumbens (NAc), thus increasing  
6 blood-brain barrier (BBB) permeability and promoting depression-like behaviors. However, the  
7 molecular mechanism underlying the reduction in tight junction protein, particularly in the NAc, is  
8 largely uncharacterized.

9 **Methods:** We performed comparative metabolomic profiling of the nucleus accumbens, prefrontal  
10 cortex and hippocampus of social defeat-stressed mice to identify the molecular events that  
11 mediate BBB breakdown.

12 **Results:** We identified the levels of cAMP were specifically reduced in the NAc and positively  
13 correlated with the degree of social avoidance. Replenishing cAMP in the NAc was sufficient to  
14 improve BBB integrity and depression-like behaviors. We further found that cAMP levels were  
15 markedly decreased in neurons of the NAc, rather than in endothelial cells, astrocytes or microglia.  
16 RNA-Seq data showed that adenylyl cyclase 5 (Adcy5), an enzyme responsible for the synthesis  
17 of cAMP from ATP, was predominantly expressed in the NAc and exclusively resided in neurons.  
18 Endogenous modulation of cAMP synthesis in neurons through the knockdown of Adcy5 in the  
19 NAc regulated the sensitivity to social stress. Moreover, deficient neuronal cAMP production in  
20 the NAc decreased the expression of reelin, while supplementary injection of exogenous reelin  
21 into the NAc promoted BBB integrity and ameliorated depression-like behaviors.

22 **Conclusions:** Chronic social stress diminished cAMP synthesis in neurons, thus damaging BBB

1 integrity in the NAc and promoting stress vulnerability. These results characterize  
2 neuron-produced cAMP in the NAc as a biological mechanism of neurovascular pathology in  
3 social stress.

4

5 Keywords: social stress, metabolomics, nucleus accumbens (NAc), cAMP, blood-brain barrier  
6 (BBB), stress vulnerability

7

8

9

## 10 **Introduction**

11 Major depression is a common illness, with a lifetime risk of approximately 15~18%  
12 worldwide(1, 2). Shortcomings in the effectiveness of now-available antidepressant treatments  
13 have resulted in an enormous public health burden of depression(3, 4). Cumulative studies have  
14 outlined that inflammation appears to precede depression(5), and patients diagnosed with major  
15 depressive disorder (MDD) have higher levels of circulating pro-inflammatory cytokines(6, 7).  
16 Peripheral inflammation can activate microglia and immune cells embedded in the brain, thereby  
17 disrupting the plasticity and development of nerve cells(6, 8).

18 Emerging evidence suggests that peripheral monocytes cannot infiltrate the brain parenchyma  
19 of stress-susceptible mice, but accumulate at the endothelial surface and perivascular space,  
20 thereby resulting in vascular remodeling(9-12). A previous study outlined that social stress  
21 specifically alters blood-brain barrier (BBB) integrity in the nucleus accumbens (NAc) through  
22 downregulation of endothelial tight junction protein Cldn5(11). NAc is a crucial region within the

1 brain's reward circuitry(13, 14). Increased permeability of the BBB tends to appear at the NAc,  
2 indicating a more complicated mechanism that mediates neurovascular remodeling in stressed  
3 mice. However, the molecular mechanism under which endothelial tight junction protein is  
4 particularly downregulated in the NAc of stress-susceptible mice is so far unclear. Herein, we  
5 identified a specific reduction in the levels of cAMP in the NAc through comparative  
6 metabolomics analysis. The data suggested that neuronal cAMP in the NAc orchestrates BBB  
7 integrity and the resultant effects on depression-like behaviors through a reelin mediated  
8 extracellular matrix network. Our study thus extends the understanding of BBB breakdown in the  
9 NAc associated with chronic social stress, providing a new perspective for understanding  
10 neurovascular pathology in depression.

11

12

### 13 **Methods and Materials**

14

#### 15 **Animals**

16 **Male C57BL/6J mice (7-8 weeks of age) and male CD-1 mice (retired breeders, 4-6 months of age)**  
17 were used (Vital River Laboratories, Beijing, China). C57BL/6J mice were housed five per cage  
18 and CD-1 mice were singly housed under standard laboratory conditions with a 12-h light-dark  
19 cycle (lights were on from 7 a.m. to 7 p.m.), temperature ( $24\pm 1^{\circ}\text{C}$ ), food and water *ad libitum*. The  
20 mice were acclimated for a minimum of 7 d prior to experiment initiation. All animal experimental  
21 procedures were approved by the Animal Ethics Committee of China Pharmaceutical University  
22 and performed in accordance with the US National Institutes of Health Guidelines.

1

## 2 **Social defeat stress model**

### 3 **Chronic social defeat stress (CSDS)**

4

5 Ten-day chronic social defeat stress was performed as previously described(15). Briefly,  
6 aggressive CD-1 mice were selected by a 3-d screening process. During the 10-d defeat period,  
7 each intruder C57BL/6J mouse was exposed to a novel resident aggressive CD-1 mouse to be  
8 physically defeated for 10 min on 10 consecutive days. Then, the intruder was transferred to  
9 one-half of the cage (the opposite compartment separated by a perforated Plexiglas divider) to  
10 maintain sensory contact for the remainder of the 24-h period. Control C57BL/6J mice were pair  
11 housed in the home cage, one mouse per side of the perforated Plexiglas divider to avoid physical  
12 contact with their cage mates. Social interaction test was performed 24 h after the last defeat.

### 13 **Microdefeat stress**

14 The microdefeat protocol is a variant of the chronic social defeat protocol, as previously  
15 described(16). Experimental C57BL/6J mice were subjected to subthreshold levels of social defeat  
16 to evaluate the increased susceptibility to stress. Three consecutive 5-min defeat sessions were  
17 conducted within the same day with 15 min of rest between each bout. Social interaction was  
18 tested 24 h later.

19

## 20 **Behavioral assays**

### 21 **Social interaction test**

22 Social interaction (SI) test was performed as previously described with minor modifications(15).

1 First, the mice were adapted to the environment of the experimental room for more than 30 min.  
2 Then, the mice were placed in an open-field arena (40 cm× 40 cm× 40 cm), containing a small  
3 wire animal cage placed on one side. Social avoidance behavior was measured according to a  
4 two-stage social interaction test, recorded with a Logitech camera (C170) and analyzed by  
5 Any-maze software (Global Biotech Inc.). In the first stage, the behavior of C57BL/6J mice was  
6 recorded for 150 s, without a CD-1 aggressor in the interaction zone. At the end of this stage, the  
7 mouse was removed for 30 s, and the arena was cleaned. In the second stage, the same metrics  
8 were measured, with a novel CD-1 aggressor in the interaction zone. The social interaction ratio  
9 (SI ratio) was calculated by dividing the time spent in the interaction zone with the target by the  
10 time spent in the interaction zone without the target. Mice were deemed to be susceptible (SI ratio  
11 <1) or resilient (SI ratio >1).

#### 12 **Forced swim test (FST)**

13 Experimental mice were singly placed in a 5-L glass beaker (17.8 cm diameter, 27.5 cm height)  
14 containing 18-20 cm of water (23-25 °C) and videotaped for 6 min under normal light(17). The  
15 immobility time was measured for the last 4 min by two colleagues who were blinded to the  
16 experiment. A stop watch was used to calculate the active behavior (swimming and climbing), and  
17 the rest immobility time was analyzed.

#### 18 **Tail suspension test (TST)**

19 Experimental mice were individually suspended by the tails with adhesive tape in the tail  
20 suspension box, roughly 20 cm above the ground and blinded to each other. The animals were  
21 video-recorded for 6 min, and the time of immobility was quantitated over the last 4 min by two  
22 colleagues who were blinded to the experiment. Immobility was defined as no movement at all or

1 only minor limb movements.

## 2 **Sucrose preference test (SPT)**

3 The sucrose preference test was performed during the dark period, started with an adaptive  
4 phase to water and 1% sucrose for 24 h, during which the two bottles were switched every 12 h.  
5 After water deprivation for 12 h, all bottles were weighed and mice were individually housed with  
6 free access to water or 1% sucrose for a 12-h period. The bottles were then reweighed and the  
7 sucrose preference was calculated by dividing the amount of sucrose consumed by the total liquid  
8 weight consumed.

## 10 **Metabolomics study and multivariate statistical analysis**

11 The metabolomics study of the samples, e.g., brain tissues of the mice, was performed based on  
12 a well-developed GC-MS and LC-MS/MS platform, as previously reported(18-20). The details  
13 regarding sample preparation, derivatization, chromatography conditions and detection of the  
14 analytes for GC-MS and LC-MS/MS can be found in the supplemental information.

## 16 ***In vivo* exposure of meglumine cAMP in mice and drug treatments**

17 To evaluate the plasma and brain (NAc) distribution of meglumine cAMP, mice were  
18 intraperitoneally administered meglumine cAMP (10 mg/kg, dissolved in 0.9% saline) or 0.9%  
19 saline as a control. 100  $\mu$ L blood samples (collected from ophthalmic veins) and NAc tissues were  
20 collected at 30 min, 1, 2, 4, 6, 12 and 24 h. The collected blood samples were centrifuged at 8,000  
21 g for 5 min. The supernatant and the brain tissues were immediately stored at -70 °C until further  
22 analysis. The concentration of cAMP in the plasma and NAc tissue was determined using a cAMP



1 ELISA kit.

2 To examine the prophylactic effect of meglumine cyclic adenylylate (M-cAMP) supplementation,  
3 M-cAMP (10 mg/kg, dissolved in 0.9% saline) was intraperitoneally (i.p.) administered 2 h prior  
4 to the social defeat, once daily for 10 d during the CSDS. Behavioral tests were performed 24 h  
5 after the last M-cAMP administration. To examine the effect of NAc supplement of M-cAMP,  
6 selected susceptible mice were anesthetized with isoflurane (4% for induction, followed by 1% for  
7 maintenance, MSS Int. Ltd, UK) and positioned in a stereotaxic instrument (RWD Life Science).  
8 Bilateral NAc (bregma coordinates: anteroposterior +1.6 mm, mediolateral  $\pm$  1.4 mm,  
9 dorsoventral -4.4 mm) were infused with 0.5  $\mu$ L M-cAMP or saline at a rate of 0.1  $\mu$ L/min(11,  
10 21). Social interaction tests were performed on day 1, 4 or 7 after injection. TST and FST were  
11 respectively performed on day 8 and 9 after injection.

12 For reelin supplementation, recombinant mouse reelin (0.5  $\mu$ g/ $\mu$ L, 0.75  $\mu$ L, 3820-MR-025,  
13 R&D Systems)(22, 23) or vehicle (1% bovine serum albumin in sterile PBS) was stereotactically  
14 injected into the NAc bilaterally, using the above mentioned coordinates for the NAc. Microdefeat  
15 stress and behavioral tests were performed three days after the injection.

16 For the fluoxetine treatment, unstressed controls and selected susceptible mice were randomly  
17 divided into either vehicle or fluoxetine (10 mg/kg, daily i.p. administration) treatment groups.  
18 NAc tissues were collected either on day 5 or 35 for western blot analysis.

19

## 20 **Stereotaxic surgery and viral gene transfer**

21 For bilateral stereotaxic injection, mice were anesthetized with isoflurane and placed in a fixed  
22 position in the small-animal stereotaxic instrument (RWD Life Science). The coordinates of NAc

1 were located according to the bregma: anteroposterior +1.6 mm, mediolateral  $\pm 1.4$  mm, and  
2 dorsoventral -4.4 mm. Each side of NAc was infused with 0.5  $\mu\text{L}$  virus (AAV2/9-shRNA or  
3 AAV2/9-shRNA-Adcy5,  $2.38 \times 10^{12}$  v.g./mL,  $2.02 \times 10^{12}$  v.g./mL, respectively) at a rate of 0.1  
4  $\mu\text{L}/\text{min}$ , and the needle was retained for 10 min before removal. The mice were post hoc screened  
5 according to the expression of GFP.

6

### 7 **Immunofluorescence**

8 Mice were transcardially perfused with cold saline, and brains were postfixed and sectioned.  
9 More details are described in the supplemental information.

10

### 11 **Capillary extraction and ELISA measurement**

12 The blood vessels of the NAc were isolated as previously described (11, 24), and more details  
13 are described in the supplemental information.

14

### 15 **Transmission electron microscopy (TEM)**

16 TEM experiments were performed as described in the supplemental information.

17

### 18 **RNA sequencing, real-time quantitative PCR, and western blot analysis**

19 The complete details of RNA sequencing, real-time quantitative PCR, and western blot analysis  
20 are provided in the supplemental information.

21

### 22 **Statistical analysis**

1 Behavioral experiments and analyses were performed by colleagues blinded to the treatment  
2 assignments. Mice were randomly grouped and the number of replicates is indicated by the dots in  
3 the figures or in the figure legends. The values that were greater than 2 s.d. from the mean were  
4 identified as outliers and excluded from further statistics. All experimental data are presented as  
5 the mean  $\pm$  s.e.m. Statistical analyses were performed using GraphPad Prism 7.0 software.  
6 Differences between two groups were examined by the unpaired Student's *t*-test. Differences  
7 among multiple groups were evaluated using one-way ANOVA or two-way ANOVA followed by  
8 Bonferroni's *post hoc* comparisons.  $P < 0.05$  was considered statistically significant. Correlations  
9 were assessed with Pearson's correlation coefficient or Spearman correlation coefficient. The  
10 incidence of depression susceptibility was analyzed with Fisher's exact test. Heatmaps were  
11 performed using R and visual representation of mean values. A summary of the statistical analyses  
12 is listed in Table S1.

#### 14 **Data availability**

15 The datasets are available from the corresponding author upon request.

#### 17 **Results**

##### 18 **Social stress vulnerability is associated with reduced cAMP in the NAc**

19 Reduced expression of the tight junction protein Cldn5 in the NAc was shown to alter BBB  
20 integrity, thus promoting depression-like behaviors; however, Cldn5 expression and BBB integrity  
21 were shown to remain unchanged in the prefrontal cortex (PFC) and hippocampus (Hip.)(11).  
22 Consistent with this finding, we observed an intense and persistent increase of IL-6 levels in the

1 plasma and NAc of the susceptible (SS) mice 24 h after the social interaction (SI) test (Figure  
2 S1A-S1E), and further verified the downregulation of Cldn5 in the NAc (Figure S1F). No  
3 difference was observed of IL-1 $\beta$  and TNF- $\alpha$  levels in the brain despite a slight increase in plasma  
4 (Figure S1E). To characterize the molecular mechanisms that underlie the region-specific  
5 breakdown of BBB integrity in the NAc, we performed comparative metabolomic profiling of the  
6 three candidate sites (NAc, PFC and Hip.) involving impaired functions in MDD of unstressed  
7 control (CTRL), susceptible (SS) and resilient (RES) mice after a 10-d chronic social defeat stress  
8 (CSDS) (Figure 1A-1D). Metabolome analyses can be employed to discover pathogenesis  
9 biomarkers for diseases, such as cardiovascular disease(25), dyslipidemia(26), cancer(27), and  
10 Alzheimer's disease(28). Here, a total of 255 endogenous metabolites were detected by GC-MS  
11 and LC-MS/MS. Figure 1E shows the differential metabolites specifically varied in the NAc of SS  
12 mice compared to those in the CTRL and RES mice, while no significant difference of these  
13 metabolites was observed in the PFC or hippocampus. Figure 1F is a heatmap showing the  
14 correlation of the concentration of these metabolites with the SI ratio. Notably, the level of cAMP  
15 was significantly lower in the NAc of SS mice (Figure 1G) and positively correlated with the  
16 degree of social avoidance (Figure 1H). Similar data were obtained using an ELISA-based  
17 approach (Figure 1I). Furthermore, the cAMP concentration in the NAc was higher than that in the  
18 PFC or hippocampus. Together, these results suggest that the distinctive change in cAMP in the  
19 NAc may underlie depression-like behaviors.

20

### 21 **cAMP supplementation improves blood-brain barrier and depressive-like behaviors**

22 To test whether replenishing cAMP could improve depressive behaviors, we examined the effects

1 of meglumine cyclic adenylylate (M-cAMP) pretreatment (Figure 2A-2E). M-cAMP is a clinical  
2 drug used for cardiovascular treatment that can readily cross cell membranes. To evaluate the  
3 kinetics of daily i.p. M-cAMP treatment, we quantified the plasma levels and brain distribution of  
4 cAMP after a single dose of 10 mg/kg M-cAMP. We verified that the levels of cAMP in  
5 circulation and the NAc were sharply increased up to 6 h post injection, but not 12 and 24 h  
6 (Figure 2F-2G). In the social interaction test, M-cAMP-pretreated mice exhibited higher SI ratios  
7 (Figure 2B) and a lower incidence of depression susceptible than vehicle-treated mice after the  
8 10-d social defeat stress (Figure 2C, Fisher's exact test,  $P=0.0033$ ). In the TST and FST, M-cAMP  
9 pretreatment significantly reduced the immobility time (Figure 2D-2E), indicating that cAMP had  
10 antidepressant-like effects. The spatial overlap between the BBB leakage and decreased cAMP  
11 levels specific in the NAc implied an association, but not necessarily a cause-and-effect. Herein, to  
12 examine whether M-cAMP pretreatment affected the BBB permeability of the NAc, we detected  
13 Cldn5 expression by immunofluorescence colocalization with CD31 and BBB leakiness by the  
14 infiltration of Evans Blue (EB). Pretreatment with M-cAMP rescued Cldn5 expression (Figure  
15 2H-2I), and M-cAMP-pretreated mice displayed no distinct infiltration of EB in the NAc (Figure  
16 2J).

17 To further confirm the effect of cAMP in the NAc on depressive behaviors, we performed  
18 stereotactic intra-NAc injection of M-cAMP or saline-vehicle to the susceptible mice (Figure 2K).  
19 A single intra-NAc injection of M-cAMP gradually improved the social avoidance behavior within  
20 one week (Figure 2L), and significantly reduced the immobile time in the TST and FST assays  
21 (Figure 2M-2N). These results support the hypothesis that replenishing cAMP in the NAc  
22 facilitates BBB integrity and induces antidepressant-like effects.

1

2 **cAMP is markedly decreased in neurons of the NAc in stress-susceptible mice**3 cAMP is thought to promote the functions of tight junctions in endothelial cells *in vitro*(29, 30).

4 As a second messenger, the synthesis and functioning of cAMP are restricted within the cell. To

5 determine whether cAMP was decreased in blood vessels, thus mediating the downregulation of

6 Cldn5, we next enriched the blood vessels of the NAc using a brain capillary extraction method.

7 After capillary extraction, the vascular sections formed a pellet in the bottom, with myelin debris

8 floating at the surface, and both of which were separately collected for measuring cAMP

9 concentration. Unexpectedly, no difference was detected for cAMP levels in NAc blood vessels of

10 SS mice when compared to control and RES mice (Figure 3A). However, the cAMP level in the

11 myelin debris of SS mice was significantly attenuated, and the cAMP level in the myelin debris of

12 the NAc was remarkably correlated with the degree of social avoidance (Figure 3B). To further

13 determine the cell type that mediates the decreasing of cAMP, we performed the

14 immune-colocalization analysis. Immunostaining revealed that cAMP was precisely colocalized

15 with the neurons and markedly decreased in neurons in the NAc, but not in astrocytes or microglia

16 (Figure 3C-3E).

17 cAMP is synthesized from ATP by adenylate cyclase located on the inner side of the plasma

18 membrane and degraded to AMP by phosphodiesterase enzymes. Along with the reduction of

19 cAMP (Figure S2A), the metabolomic profiling also revealed that ATP was accumulated (Figure

20 S2B), while AMP was constant in the NAc of SS mice (Figure S2C), indicating that the activity of

21 adenylate cyclase was disturbed in the NAc. Our RNA-sequencing (RNA-seq) data displayed that

22 the NAc region predominantly expressed adenylate cyclase 5 (*Adcy5*) among the 10 subtypes

1 (Figure 3F). Moreover, a single cell RNA sequencing of the mouse nervous system mapped that  
2 *Adcy5* was mainly expressed in the medium spiny neurons of the NAc (Figure S2D)(31). Adenylyl  
3 cyclase is activated by coupling with G $\alpha$ , especially the G $\alpha$  located in nonlipid raft region,  
4 where it is more facile to activate adenylyl cyclase(32, 33). We next explored the protein  
5 expression of *Adcy5* and G $\alpha$  both in lipid raft region and non-raft region, using flotillin-1 as a  
6 neuron specific lipid raft marker. No difference of protein expression of *Adcy5* was observed in  
7 either the lipid raft or non-raft region, however, G $\alpha$  expression was markedly dampened in  
8 non-raft region of SS mice, mediating a reduced ability of *Adcy5* to convert ATP to cAMP in the  
9 NAc (Figure 3G-3H).

10 Taken together, these results demonstrated that the lessened cAMP in the NAc of SS mice was  
11 occurred in the neurons, and was mediated by less G $\alpha$  translocation from lipid rafts to non-raft  
12 region.

#### 14 **Behavioral changes following endogenous modulation of neuronal cAMP synthesis in the** 15 **NAc**

16 To further confirm the causal role of cAMP in social stress induced depressive behaviors, we  
17 conducted conditional knockdown of *Adcy5* to modulate the neuronal cAMP concentration in the

18 NAc *in vivo*. A doxycycline-inducible adeno-associated virus (AAV 2/9 serotype) targeting shRNA  
19 transcripts encoding *Adcy5* allows for reversible modulation of cAMP in the NAc (Figure S3A).

20 We confirmed the downregulation of *Adcy5* mRNA and protein, as well as lower cAMP levels in  
21 the NAc (Figure S3B-S3G). No difference was found for other subtypes of adenylate cyclase.

22 After 2 weeks of recovery followed by 2 weeks of treatment with doxycycline, the microdefeat, a

1 subthreshold level of social defeat, was employed to investigate the pro-susceptibility effect of  
2 reductive cAMP in the NAc (Figure 4A). We consistently observed that the reduction in cAMP  
3 alone had no effect on the behaviors, however, the cAMP reduction in combination with exposure  
4 to microdefeat markedly induced social avoidance behavior and anhedonia (Figure 4B-4C, Figure  
5 S4A-S4B). Notably, withdrawal of doxycycline in the following 5 days, allowing recovery of  
6 *Adcy5* expression (Figure S3H-S3K), significantly reversed the depressive behaviors (Figure  
7 4D-4F, Figure S4C). Moreover, another cohort of mice conducted replenishing cAMP also  
8 displayed insusceptible to the microdefeat (Figure 4G-4L, Figure S4D-S4F). Together, these  
9 results suggest that neuronal cAMP deficiency in the NAc enhanced stress susceptibility in mice.

10

#### 11 **Reelin mediates the antidepressant-like effects of cAMP**

12 The RNA-seq analysis indicated that the extracellular matrix (ECM)-receptor interaction  
13 pathway was markedly disturbed in the NAc of SS mice compared to that in CTRL mice after a  
14 10-d CSDS (Figure S5A). Among the genes in this pathway, the gene expression of collagen IV  
15 (*Col IV*), which is the major structural component of basement membranes, was significantly  
16 reduced. Colocalization of Col IV with CD31 and transmission electron microscopy showed that  
17 the basement membrane of the BBB, not just endothelial tight junctions, was compromised in the  
18 NAc (Figure S5B-S5C), while replenishing cAMP improved the integrity of the basement  
19 membrane in the NAc (Figure S5D). Reelin (Reln), a secreted glycoprotein mainly expressed by  
20 GABAergic interneurons, has been demonstrated to play a vital role in the extracellular matrix  
21 network that promotes BBB development through astrocytic endfeet attachment to the vasculature  
22 (34, 35). Notably, the expression of *Reelin* in the cerebrum was spatially coincided with *Adcy5*,



1 according to the Allen Brain Atlas (Figure S6). We therefore further examined whether the cAMP  
2 level in the NAc potentially impacted the reelin signal. Interestingly, we found that the mRNA and  
3 protein expression of reelin was much lower in AAV-shRNA-Adcy5 mice than in AAV-shRNA  
4 mice (Figure 5A-5B), suggesting a crucial role of reelin in the cAMP deficiency mediated  
5 pro-susceptibility. Peripheral IL-6 (~21 kDa) was reported to be able to penetrate the NAc  
6 parenchyma of SS mice. We concomitantly found that the recombinant biotinylated IL-6 was  
7 detectable in the NAc of stressed AAV-shRNA-Adcy5 mice, but not stressed AAV-shRNA mice  
8 (Figure 5C). Notably, intra-NAc injection of reelin to the AAV-shRNA-Adcy5 mice was sufficient  
9 to ameliorate social avoidance behavior in SI test (Figure 5D-5F) and anhedonia behavior in the  
10 sucrose preference test (Figure 5G), as well as the infiltration of biotinylated IL-6 in the NAc  
11 (Figure 5H). We also evaluated the effect of acute (5 d) and chronic (35 d) fluoxetine treatment on  
12 the reelin signal in the NAc of SS mice. Chronic fluoxetine treatment (Figure 5J) but not acute  
13 fluoxetine treatment (Figure 5I) was sufficient to recuperate reelin protein expression in the NAc.

14

## 15 **Discussion**

16

17 In general, our data suggest that neuronal cAMP in the NAc orchestrates BBB integrity and  
18 stress vulnerability. cAMP was reported to be diminished globally in the brains of patients with  
19 major depressive disorder (MDD) and was upregulated by antidepressant treatment(36, 37).  
20 However, our current knowledge of cAMP in MDD, especially in some particular brain regions, is  
21 fairly limited (38). Our findings complement previous evidence suggesting that replenishing  
22 cAMP can ameliorate the BBB integrity of the NAc of stress-susceptible mice. BBB leakiness in

1 depression was first brought up several decades ago(39), and the most recent developments  
2 highlighted that the BBB integrity of stress-susceptible mice was specifically compromised in the  
3 NAc(11). Moreover, peripheral monocytes cannot infiltrate the brain parenchyma, but accumulate  
4 at the endothelial surface and perivascular space(9, 10). Small molecules and proteins up to ~69  
5 kDa have been reported to pass through the leaky BBB of stress-susceptible mice(11). The  
6 extracellular matrix signal, also identified to be related to depression by some reports(40-43), has  
7 been widely studied for its promotion of neurogenesis(44), modulation of synaptic plasticity(45),  
8 and relation to cognitive deficits(46). Our work uncovers the molecular mechanism by which  
9 neuronal cAMP may regulate the BBB integrity of the NAc via reelin-mediated extracellular  
10 matrix signals.

11 A previous study focusing on the cAMP homeostasis regulated by the downstream  
12 phosphodiesterase-4 (PDE4) pathway also confirmed the pivotal role of cAMP in NAc mediated  
13 depression-like behaviors (47). Given that the present metabolomic profiling results supported that  
14 the upstream but not downstream of cAMP synthesis was disturbed in the NAc, we thus addressed  
15 the *Adcy5* mediated synthesis process of cAMP in the NAc. To our knowledge, an *in vitro* study  
16 showed that ketamine produced antidepressant effect by increasing cAMP in glial cells and  
17 astrocytes, rather than targeting NMDA receptors (48). This hints the crucial role of cAMP in  
18 depression and the different effects of cAMP in particular cell types. Considering that *Adcy5* is  
19 predominantly expressed in neurons, but also non-neurons to a minor extent, and that the viral  
20 manipulation is not cell specific, further work is required to illuminate the specific groups of  
21 neurons in the NAc that are involved in cAMP-mediated stress vulnerability. Overall, it is  
22 plausible that replenishing cAMP in the NAc or targeting the regulation of endogenous cAMP

1 may promote the design of effective antidepressant drugs. In addition, it is interesting to note that  
2 M-cAMP is a clinical drug for cardiovascular treatments, while depression and cardiovascular  
3 disease have been found with high comorbidity in patients(49-51). A greater understanding of the  
4 mechanisms by which M-cAMP enhances vascular health in MDD may promote the augmentation  
5 of current treatment protocols.

6

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15

### 16 **Disclosures**

17 The authors report no biomedical financial interests or potential conflicts of interest.

18

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22

1

2 **Figure legends**

3

4 **Fig. 1 Social stress vulnerability is associated with reduced cAMP specifically in the NAc**

5 (A) Schematic representation of the CSDS procedure. (B) Stressed mice were divided into  
6 susceptible (SS) and resilient (RES) phenotypes according to the SI ratio; (C) Heat maps of time  
7 spent in the SI test. (D) Quantification of the immobile time in the TST. (E) Heat map visualizing  
8 the intensities of the metabolites that were particularly different in the NAc. (F) Level of the  
9 metabolites correlated with SI ratio; correlations were evaluated with Pearson's correlation  
10 coefficient or Spearman correlation coefficient, \* $P < 0.05$ , \*\* $P < 0.01$ . (G) cAMP levels in the NAc,  
11 PFC and Hip. (H) cAMP levels (detected by LC-MS/MS) in the NAc correlated with SI ratio and  
12 (I) cAMP levels (measured by ELISA) in the NAc correlated with SI ratio. Correlations were  
13 evaluated with Pearson's correlation coefficient. One-way ANOVA followed by Bonferroni's  
14 multiple comparison test, \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\*\* $P < 0.0001$ , n.s., no significance. Data  
15 represent the mean  $\pm$  s.e.m. Also see supplementary Table S1 for statistical analysis.

16

17 **Fig. 2 cAMP supplementation improves blood-brain barrier and depressive-like behaviors**

18 (A) Schematic representation of the CSDS procedure and M-cAMP treatment, intraperitoneally  
19 (i.p.) administered once daily 2 h prior to social defeat. (B) Individual SI ratio values. (C)  $2 \times 2$   
20 contingency table for correlation between social avoidance and M-cAMP treatment. The numbers  
21 in the box indicate the number of animals. Fisher's exact test,  $P = 0.0033$ . (D) Quantification of the  
22 immobile time in the TST and FST (E). The plasma (F) and NAc (G) cAMP levels after a single

1 i.p. M-cAMP treatment (n=6), two-way ANOVA followed by Bonferroni's multiple comparison  
2 test, \*\*\* $P < 0.001$ , \*\*\*\* $P < 0.0001$ , n.s., no significance. (H) Quantification of the Cldn5 expression,  
3 ratio of Cldn5<sup>+</sup> area/CD31<sup>+</sup> area. (I) Representative costaining with CD31 and Cldn5 in blood  
4 vessels. Scale bars, 20  $\mu\text{m}$ . (J) Schematic representation of the CSDS procedure and M-cAMP  
5 treatment. (K) Assessment of BBB permeability with Evans Blue dye. (L) Individual SI ratio  
6 values on different days. (M) Quantification of the immobile time in the TST and FST (N).  
7 One-way ANOVA followed by Bonferroni's multiple comparison test, \* $P < 0.05$ , \*\* $P < 0.01$ ,  
8 \*\*\* $P < 0.001$ , \*\*\*\* $P < 0.0001$ , n.s., no significance. Data represent the mean  $\pm$  s.e.m. Also see  
9 supplementary Table S1 for statistical analysis.

10

11 **Fig. 3 cAMP is markedly decreased in neurons in the NAc of stress-susceptible mice**

12 cAMP levels in vascular segments (A) and myelin debris (B) in the NAc. Right: cAMP level  
13 correlated with SI ratio. Correlations were evaluated with Pearson's correlation coefficient. (C)  
14 Co-staining of the NAc with cAMP and  $\beta$ 3-tubulin (neurons), GFAP (astrocytes), and Iba1  
15 (microglia). Scale bars, 50  $\mu\text{m}$ . Scale bars of insets: 20  $\mu\text{m}$ . Staining intensity was quantified (D),  
16 and colocalization analysis was performed (E) as shown in the right panels. (F) mRNA expression of  
17 different isoforms of adenylyl cyclase in the NAc of mice. Protein expression of Adcy5 and Gs $\alpha$  in  
18 the nonlipid raft (G) and lipid raft regions (H). n = 8 experimental replicates/group, and the  
19 statistics are shown in the lower panels. One-way ANOVA followed by Bonferroni's multiple  
20 comparison test, \* $P < 0.05$ , \*\*\* $P < 0.001$ , \*\*\*\* $P < 0.0001$ . n.s., no significance. Data represent the  
21 mean  $\pm$  s.e.m. Also see supplementary Table S1 for statistical analysis.

22

1 **Fig. 4 Behavioral changes following endogenous modulation of neuronal cAMP synthesis in**  
2 **the NAc**

3 (A) Schematic representation of the *Adcy5* rescue experiment. (B) Individual SI ratio values on  
4 day 29. (C) Quantification of the immobile time in the TST on day 30. (D) Individual SI ratio  
5 values on day 38. (E) Quantification of the immobile time in the TST on day 39. (F) Assessment  
6 of sucrose preference on day 41. (G) Schematic representation of the cAMP replenishing  
7 experiment. (H) Individual SI ratio values on day 29. (I) Quantification of the immobile time in  
8 the TST on day 30. (J) Individual SI ratio values on day 38. (K) Quantification of the immobile  
9 time in the TST on day 39. (L) Assessment of sucrose preference on day 41. Two-way ANOVA  
10 followed by Bonferroni's multiple comparison test, \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\*\* $P < 0.0001$ . Data  
11 represent the mean  $\pm$  s.e.m. Dox, doxycycline. Also see supplementary Table S1 for statistical  
12 analysis.

13  
14 **Fig. 5 Reelin mediates the antidepressant-like effects of cAMP**

15 mRNA (A) and protein (B) expression of reelin in the NAc of AAV-shRNA and  
16 AAV-shRNA-*Adcy5* mice. Scale bars, 200  $\mu$ m. Scale bars of insets: 30  $\mu$ m. Unpaired *t*-test with  
17 equal variance, \* $P < 0.05$ . (C) Colocalization of biotinylated IL-6 and CD31 in the NAc. Scale bars,  
18 20  $\mu$ m. (D) Schematic representation of the reelin replenishing experiment. (E) Individual SI ratio  
19 values of mice. (F) Heat maps of time spent in the SI test. (G) Assessment of sucrose preference.  
20 (H) Colocalization of biotinylated IL-6 and CD31 in the NAc. Scale bars, 20  $\mu$ m. (I) Protein  
21 expression of reelin after acute fluoxetine treatment. (J) Protein expression of reelin after chronic  
22 fluoxetine treatment. The statistics are shown in the bottom panels. Two-way ANOVA followed by

- 1 Bonferroni's multiple comparison test, \* $P < 0.05$ , \*\* $P < 0.01$ . n.s., no significance. Data represent
- 2 the mean  $\pm$  s.e.m. Also see supplementary Table S1 for statistical analysis.

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